



Prenatal cytogenetic diagnosis : laboratory aspects

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Goal of prenatal diagnosis

To inform couples about the risk of a birth defect or genetic disorder in their pregnancy

To provide them with informed choices on how to manage that risk (genetic counseling)

Principal indications

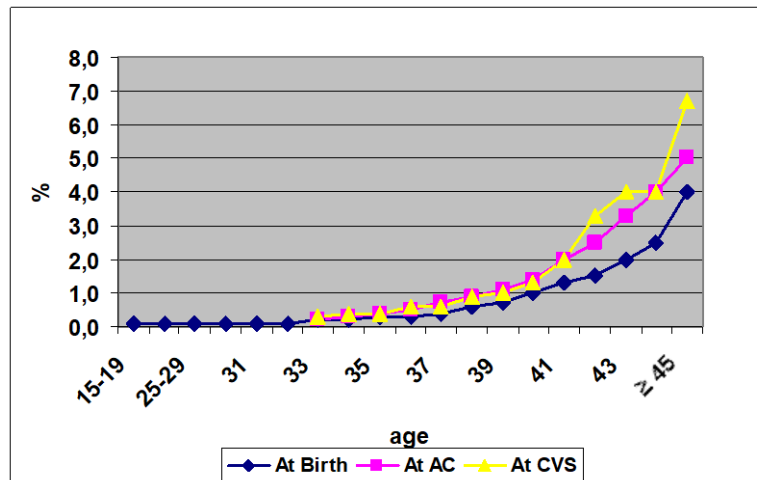
Known family history -> elevated risk for a specific genetic disorder

Ultrasound abnormalities

Advanced maternal age

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Invasive testing

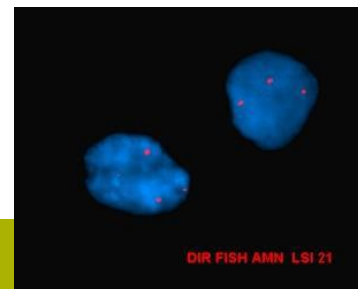
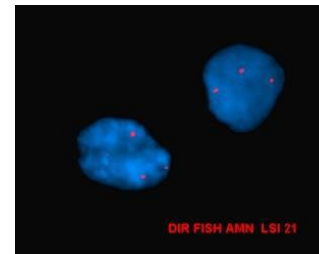
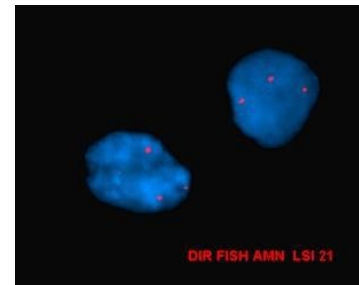
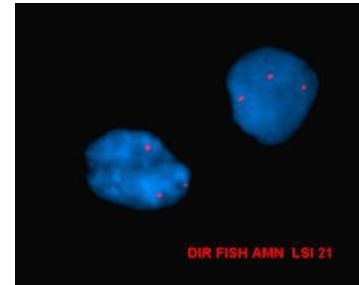
Chorionic villus sampling

Amniocentesis

- **Cordocentesis: after 20th week of gestation**
→ fetal blood
- **Preimplantation genetic diagnosis**
→ another presentation

Evolution of prenatal diagnosis

- <2010
→ All invasive samples
- 2010
→ Ultrasound anomalies
- Other indications
- 2013
→ All invasive samples



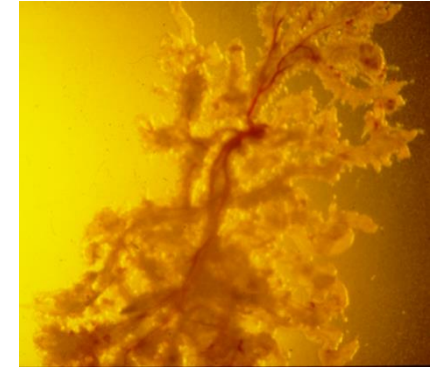
Consensus 8 genetic centers in Belgium

- **From 2013 in Belgium: for all prenatal samples = aCGH**
 - **Consensus:**
 - Use 60K arrays (or comparable resolution)
 - Always test for maternal cell contamination
 - Always obtain a parental blood sample
 - Always have at least 1 backup flask in culture
 - Testing for triploidy is done (FISH, STR, SNP array)
 - A rapid aneuploidy test is not necessary if the TAT is less than one week
 - Batching samples -> benefits for cost (lab work)

Invasive testing

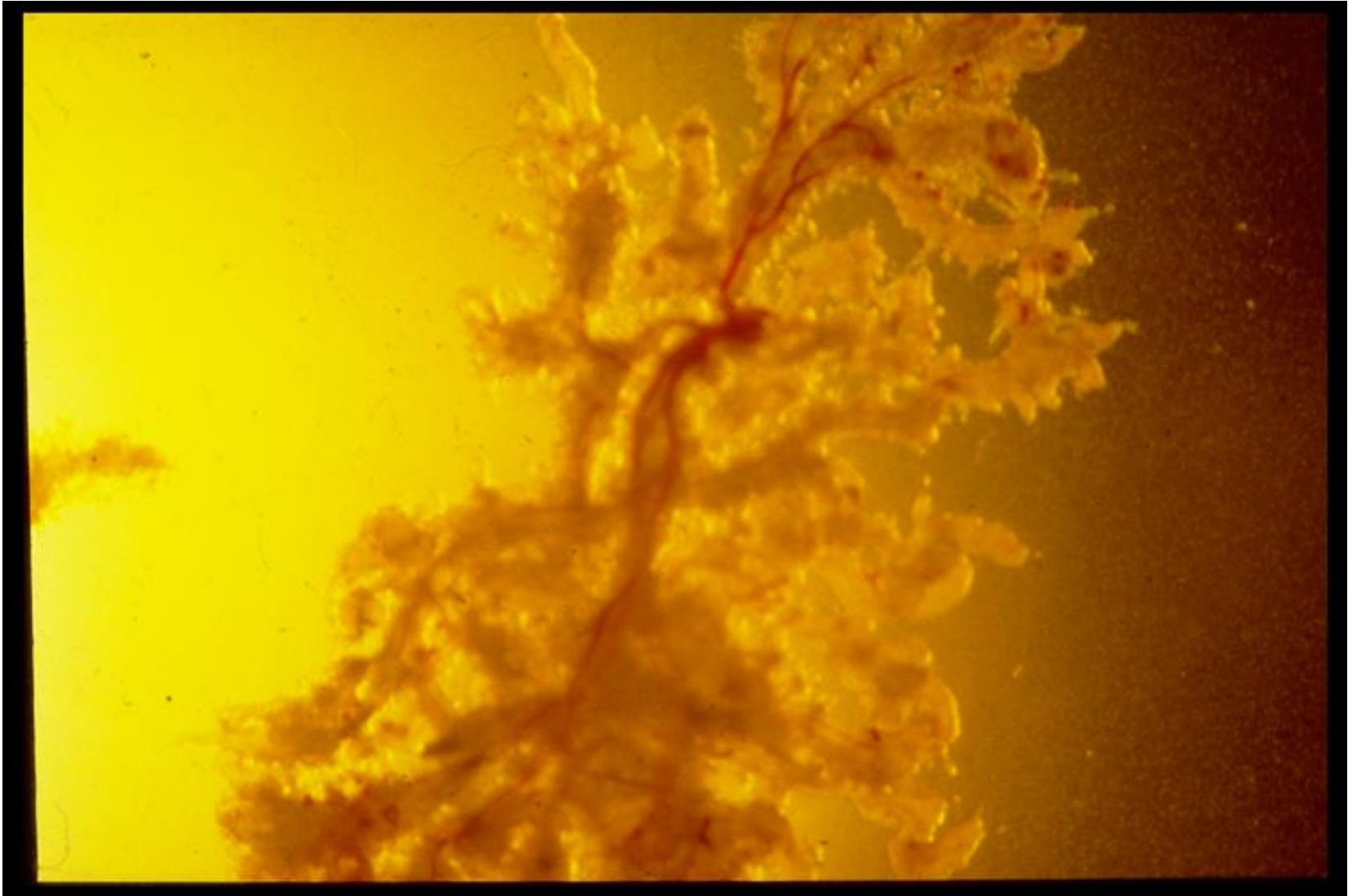
- **Chorionic villus sampling (CVS) :**
From 11 - 12th week of pregnancy

- **Amniocentesis :**
From 14 - 16th week of pregnancy



➡ **in our laboratory**

Chorionic villus sampling (CVS)



Prenatal culture room-CVS

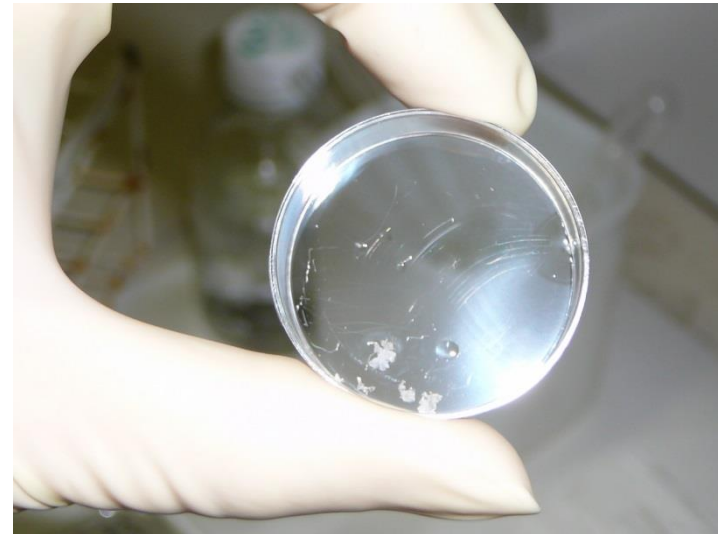
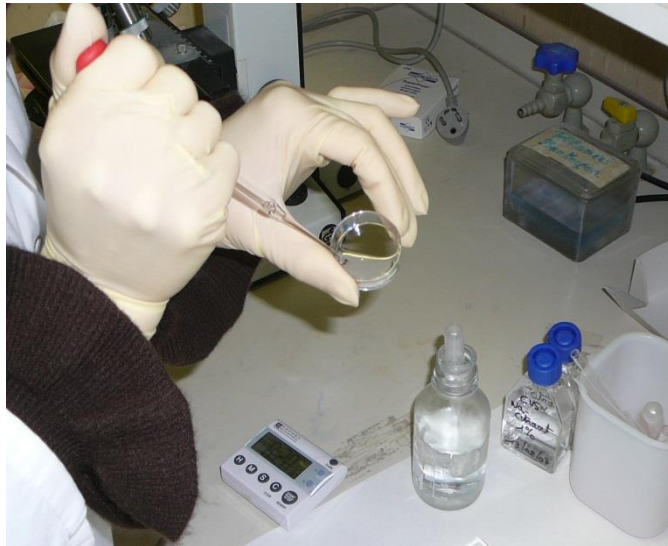


Microscopic dissection chorionic villi

1 villi (uncultured): array CGH + MCC/rapid aneuploidy (QF-PCR)

1 villi: if necessary for DNA/stock

1 villi -> short-term culture (overnight) for FISH + back-up culture (long-term, > 1 week)



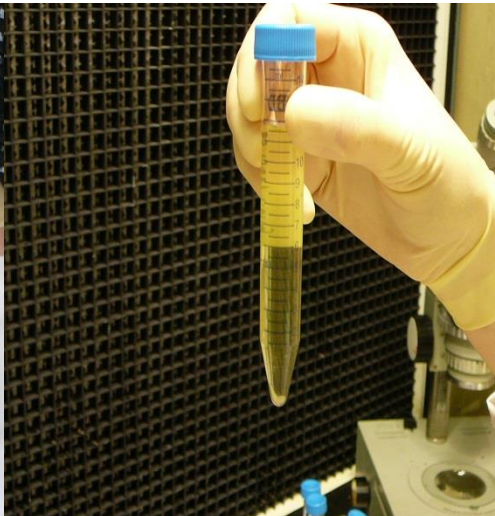
Prenatal culture room-AC



1 tube (10 ml): array CGH + MCC/rapid aneuploidy (QF-PCR)

1 tube: : if necessary for DNA/stock

1 tube: FISH (3 ml) +
back-up culture (7 ml)

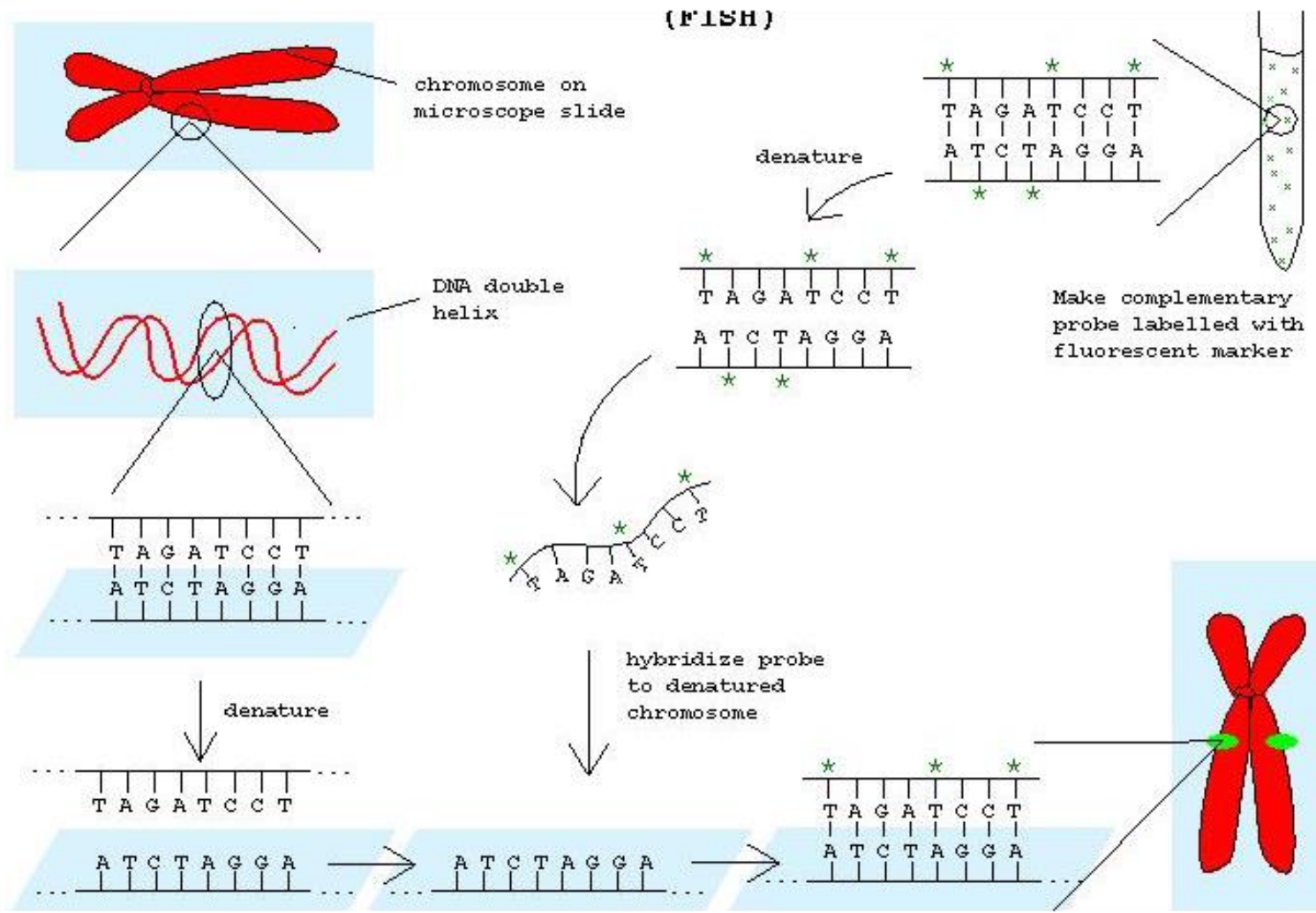


pellet



Washing

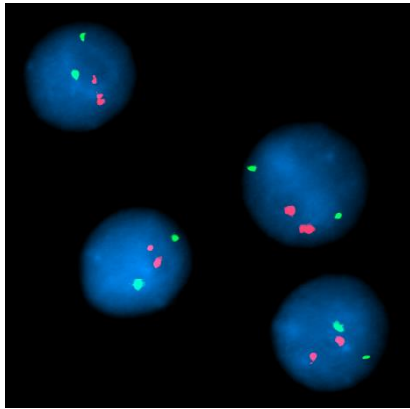
Fluorescence in situ hybridisation



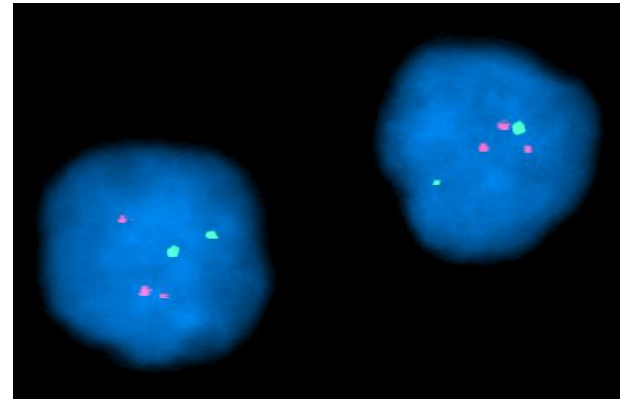
Fluorescent in situ hybridisation

Aneuploidy screening (interphase nuclei: direct test)

- X,Y,13,18,21

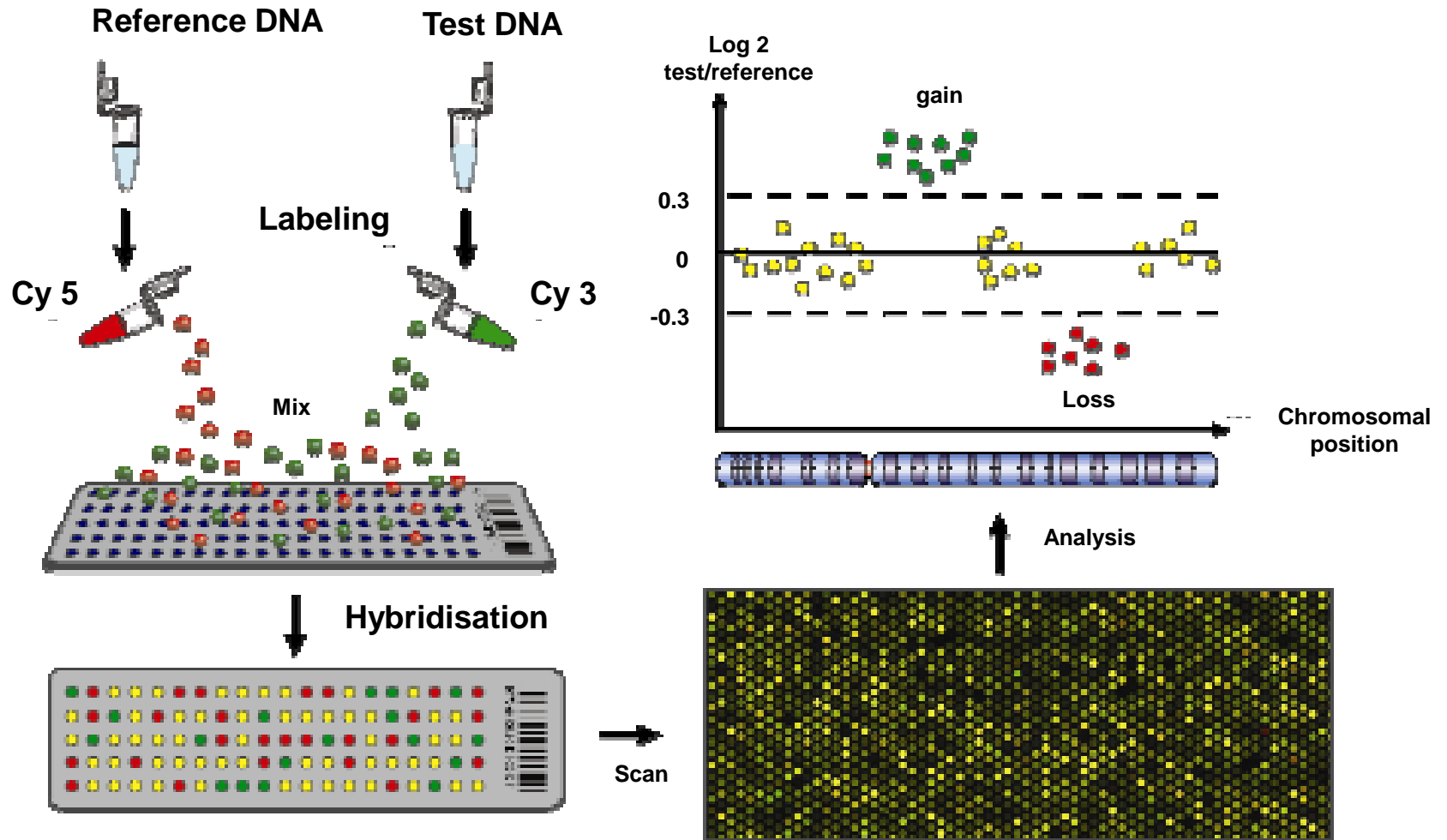


Normal result
(XX, 21)



Trisomy 21 (XX)

Array CGH-Principial



Array CGH prenatal result

- **In Belgium 2013: aCGH for all prenatal samples**
 - **consensus: to use 60K arrays (60 000 probes) or an equivalent for an average resolution of 400 kb**
 - **Additional diagnostic yield** (compared to conventional karyotyping; Shaffer et al. 2012; Wapner et al.2012):
 - **±10% in fetuses with multiple ultrasound abnormalities**
 - **± 1% in lower risk women, such as those of advanced maternal age**
 - **Drawback: introduce CNVs of uncertainty into the diagnostic interpretation**

National consensus guideline between the 8 Centres for Medical Genetics in Belgium

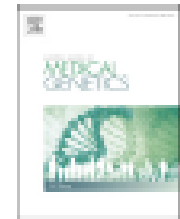
- Practical recommendation of pre- and post-counselling
 - can we expect parents to make 'on spot' decisions on what they do and do not want to know?
 - should we confront parents with questions that are unlikely to be relevant for them?
- How to interpret and report prenatal array results



Contents lists available at ScienceDirect

European Journal of Medical Genetics

journal homepage: <http://www.elsevier.com/locate/ejmg>



Review

Implementation of genomic arrays in prenatal diagnosis: The Belgian approach to meet the challenges



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Prenatal array guidelines

- **Classification of variants with regard to pathogenicity:**
 - Pathogenic
 - Benign variants without functional consequences
 - Unclassified variants (UV)

https://www.college-genetics.be/assets/recommendations/fr/guidelines/BeSHG%20prenatal%20consortium_guidelines%20prenatal%20array.pdf

Pathogenic CNV

- known to be associated with a phenotype (e.g. del22q11.2)
- resulting in a known effect on gene function and known phenotypic effect

Are communicated

Benign CNV without functional consequences

- **Is repeatedly found in the normal population and not enriched in individuals with abnormal phenotypes**

Are NOT communicated

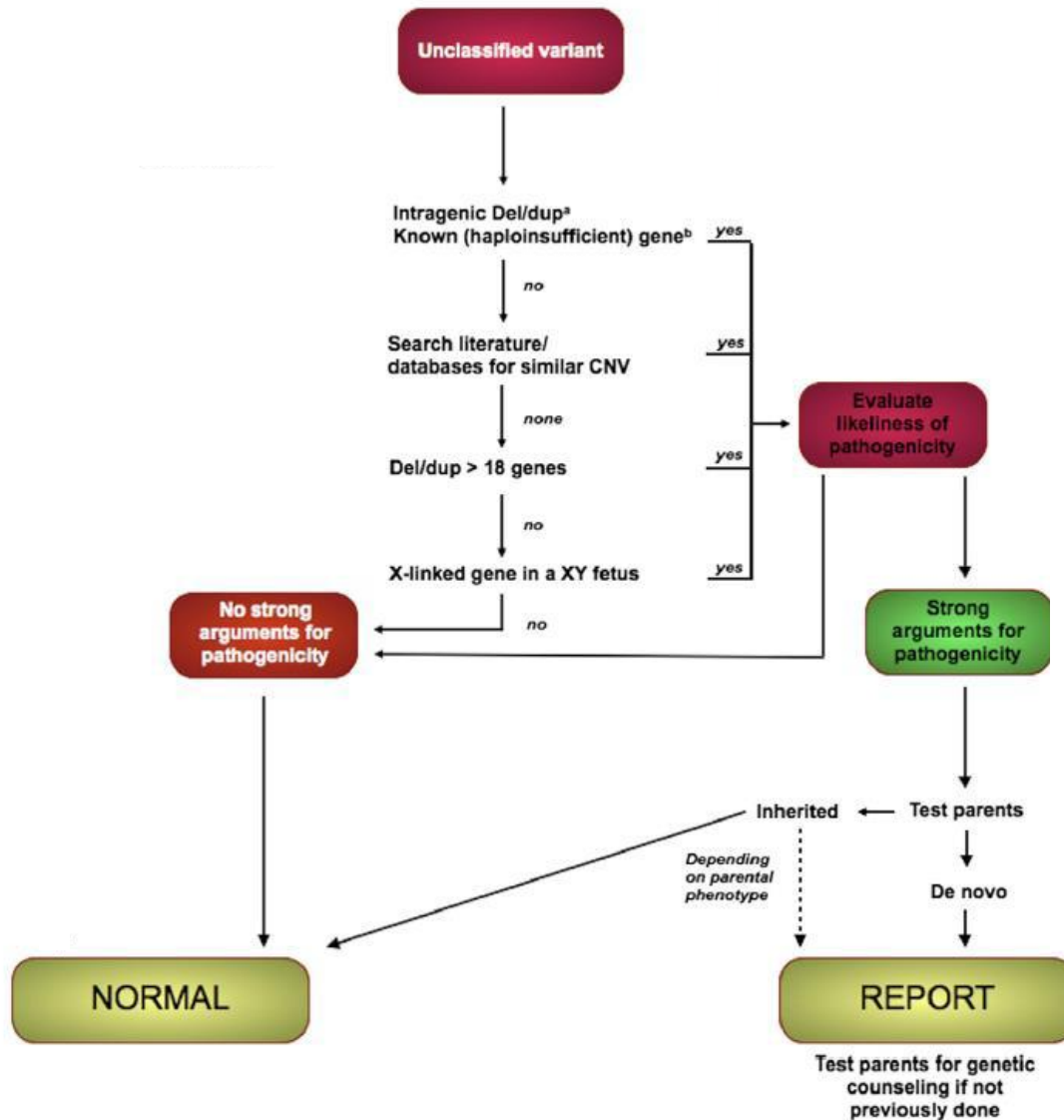
Unclassified variants (UV)

- In principle, UVs are NOT communicated and parental analysis is not performed.
 - unless one expects that this will add to the interpretation of the UV and to the decision to communicate this CNV.

Examples include CNVs with a higher degree of suspicion that they may cause a phenotype, the presence of ultrasound anomalies, family history etc.

In case of uncertainty, the ad hoc committee is consulted for advice. This is done before the final protocol is issued.

Analysis prenatal arrays



SUSCEPTIBILITY CNVs

- CNVs that are risk factors for developmental disorders

NOT communicated

- unless the risk is large enough and/or the CNV is associated with structural malformations for which ultrasound follow-up is indicated

SEE list

available on the website of the College for Genetics: <https://www.college-genetics.be/nl/voor-deprofessionele/good-practice-et-richtlijnen-voor-beroepsbeoefenaars/richtlijnen.html>.

List of susceptibility loci

| chr | start in Mb (hg19) | stop in Mb (hg19) | size in kb | CNV | gene | phenotype | morph. anomaly | return? | OMIM | update May 2017 |
|-----|--------------------|-------------------|------------|---|--------------------|--|--|-----------|--------|--|
| 1 | 146,57 | 147,39 | 820 | distal 1q21.1 dup | <i>GJA5 (CX40)</i> | ID, DD, ASD, schizophrenia | macrocephaly, CHD | YES | 612475 | YES |
| 1 | 146,57 | 147,39 | 820 | distal 1q21.1 del | <i>GJA5 (CX40)</i> | ID, DD, ASD, SZ, facial dysmorphism | microcephaly, CHD, renal and urinary tract anomalies | YES | 612474 | YES |
| 1 | 171,81 | 172,38(?) | 57 | 1q24.3 del | <i>DNM3</i> | ID | IUGR, microcephaly, brachydactyly | YES | | |
| 15 | 31,13 | 32,48 | 1350 | 15q13.3 del | <i>CHRNA7</i> | DD, ID, ASD, epilepsy, SZ | microcephaly, CHD | YES | 612001 | YES |
| 15 | 99,36 | 102,52 | 3160 | 15q26 del | <i>IGF1R</i> | MR | IUGR | YES | | YES |
| 16 | 28,74 | 28,96 | 220 | 16p11.2 distal del | <i>SH2B1</i> | obesity, DD, ID, SZ | none | YES | 613444 | YES |
| 16 | 29,59 | 30,19 | 600 | 16p11.2 proximal dup | <i>TBX6</i> | ASD, ID, DD, SZ, anorexia | microcephaly | NO YES | 614671 | moved to YES since actionable; penetrance del and dup comparable |
| 16 | 29,59 | 30,19 | 600 | 16p11.2 proximal del | <i>TBX6</i> | ID, DD, ASD, obesity, SZ, speech delay | macrocephaly, vertebra | YES | 611913 | YES |
| 17 | 34,82 | 36,21 | 1390 | 17q12 deletion syndrome RCAD (renal cysts & diabetes) | <i>TCF2</i> | facial dysmorphism, genital abnormalities, ID, DD, ASD, MODY | renal anomalies | YES | 614527 | YES |
| 22 | 19,02 | 20,29 | 1270 | 22q11.2 dup | <i>TBX1</i> | ASD, ID, DD, dysmorphic features | microcephaly, CHD | YES | 608363 | YES |
| 1 | 144,97 | 146,61 | 1640 | 1q21.1 dup | <i>HFE2</i> | DD, ASD | CHD | NO | | NO |
| 2 | 50 | 51,11 | 1110 | 2p16.3 del | <i>NRXN1</i> | ID, ASD, SZ, DD, dysmorphic features | none | NO | 614332 | NO |
| 2 | 110,87 | 110,98 | 110 | 2q13 dup | <i>NPHP1</i> | ASD, ID | none | NO | | NO |
| 3 | 197,2 | 198,84 | 1600 | 3q29 dup | | MR, DD | none | NO | | NO |
| 13 | 20,81 | 21,01 | 1200 | 13q12 dup | <i>CRYL1</i> | ? | ? | NO | | NO |
| 15 | 22,8 | 23,09 | 290 | 15q11.2 dup | <i>NIPA1</i> | DD, motor delay, speech delay, ASD | none | NO | | NO (likely benign) |
| 15 | 22,8 | 23,09 | 290 | 15q11.2 del | <i>NIPA1</i> | ID, DD, epilepsy | CHD | NO | 615656 | NO (likely benign) |
| 15 | 31,13 | 32,48 | 1350 | 15q13.3 dup | <i>CHRNA7</i> | ADHD, ID, DD, ASD | none | NO | | NO (likely benign) |
| 16 | 14,98 | 16,48 | 1500 | 16p13.11 dup | <i>MYH11</i> | ID, ASD, SZ, ADHD | aorta dilatation | NO | | NO |
| 16 | 14,98 | 16,48 | 1500 | 16p13.11 del | <i>MYH11</i> | ID, DD, ASD, epilepsy | microcephaly | NO | | NO |
| 16 | 21,94 | 22,46 | 520 | 16p12.2 dup | <i>EEF2K, CDR2</i> | ? | ? | NO | | NO (likely benign) |
| 16 | 21,94 | 22,46 | 520 | 16p12.2 del | <i>EEF2K, CDR2</i> | DD, speech delay | cranofacial and skeletal abnormalities, CHD | NO | 136570 | NO |

Incidental findings

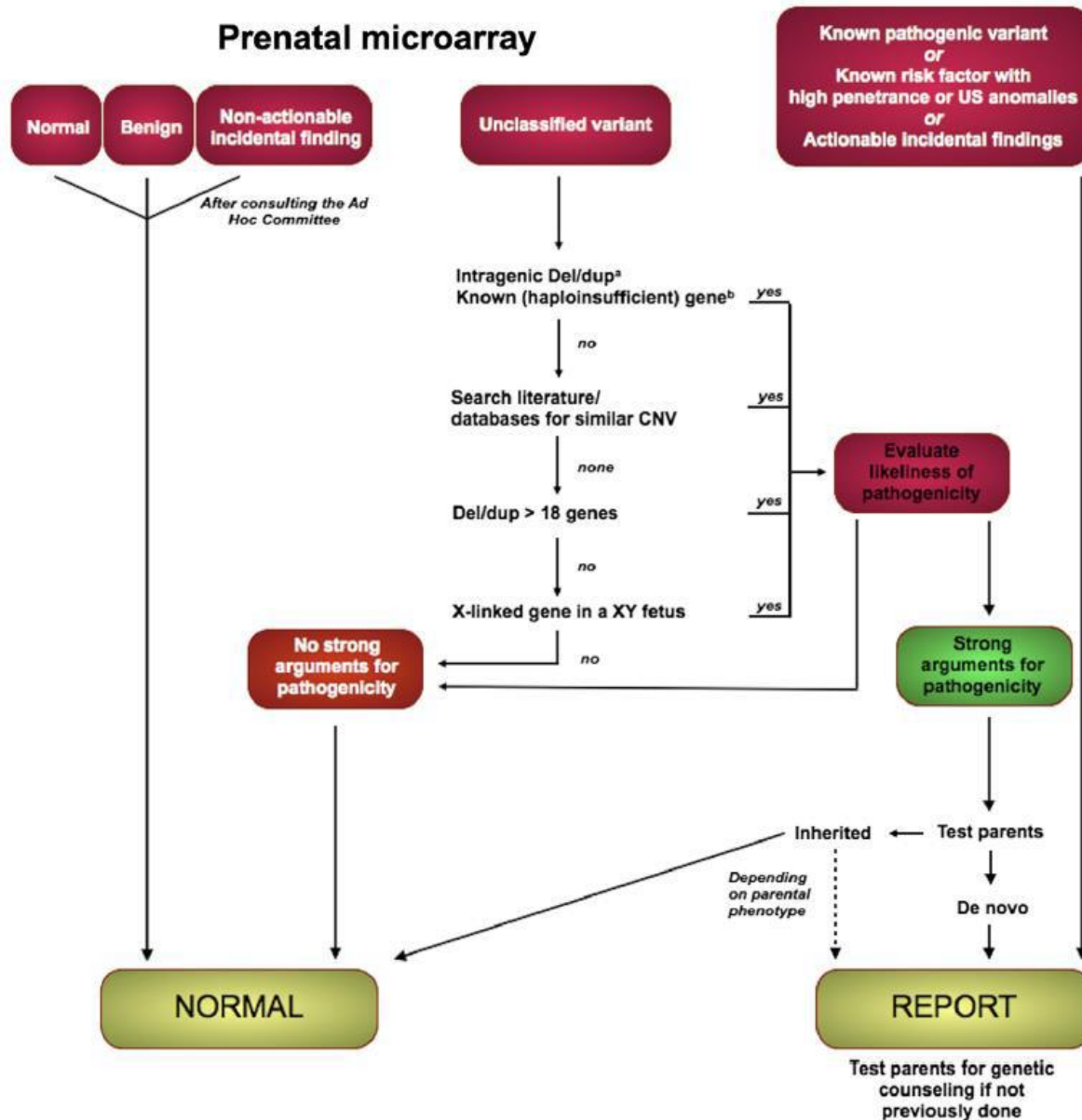
- Only highly penetrant monogenic disorders are considered, with validated evidence on the phenotype associated with the deletion or duplication

Incidental findings

Four categories are distinguished:

- **Late-onset genetic disorders with clinical utility**
 - will be communicated (typically cancer caused by the deletion of a tumor suppressor gene)
- **Late onset disease without therapeutic possibilities**
 - the decision after consulting the ad hoc committee
- **Carrier for X-linked recessive disorders**
 - will be communicated
- **Carrier for autosomal recessive disorders**
 - will not be communicated

Analysis prenatal arrays



Implementation of an *Ad Hoc* committee

- 2 clinical geneticists and 2 cytogeneticist from each center = 32 individuals
- cases are presented to the committee through e-mail
- AIM: to reach a consensus decision within 24-48h
- less subjective
- more consistent counselling in case of second opinion in another centre
- rapid learning curve on evaluation of 'difficult' CNVs

Advisory role



Clinician holds responsibility on final decision

To Do / Ongoing national guidelines

- **Regular re-evaluation to further optimize the consensus approach**
- **Address several outstanding questions**
 - **proportion of cases with unclassified variant?**
 - **% detection of causal CNVs in different indications?**
 - **% of incidental findings?**
 - **how often is parental analysis indicated?**
 - **incidence of susceptibility loci?**
 - **detection of causal CNVs postnatally?**
 - **postnatal follow-up**

Conclusion national guidelines

- **The National consensus approach solves:**
 - **technical issues (resolution, what to test for, etc..)**
 - **variation in interpretation amongst laboratories**
 - **variation of reporting**
 - **issues related to liability**

**Practical aid for those routinely using
prenatal arrays**

Conclusion national guidelines

info@college-genetics.be

Nederlands ▾

Contact

Toegang leden

Search



Richtlijnen

Onze taken

Wetgeving

Samenstelling

Nieuws

Plan voor Zeldzame Ziekten ▾

Voor de beroepsbeoefenaars ▾

Voor de patiënten ▾

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| BeSHG prenatal consortium_guidelines for fetal genome-wide sequencing (NGS) in ongoing pregnancies | → |
| BeSHG prenatal consortium_guidelines for prenatal rasopathy panel | → |
| BeSHG prenatal consortium_guidelines managing incidental findings detected by NIPT | → |
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| BeSHG prenatal consortium_table susceptibility loci | → |
| COVID19_WHO_Laboratory biosafety | → |



Vrije Universiteit Brussel

Mosaicism in prenatal diagnosis

- **Mosaicism**

- **Is difficult for making a conclusion**

- **The presence of two or more cell lines in a tissue sample**

- **Three categories**

- **Confined placental mosaicism**

- **True Constitutional fetal mosaicism**

- **Pseudomosaicism refers to an abnormality that arose during tissue culture in vitro (cultural artifact)**

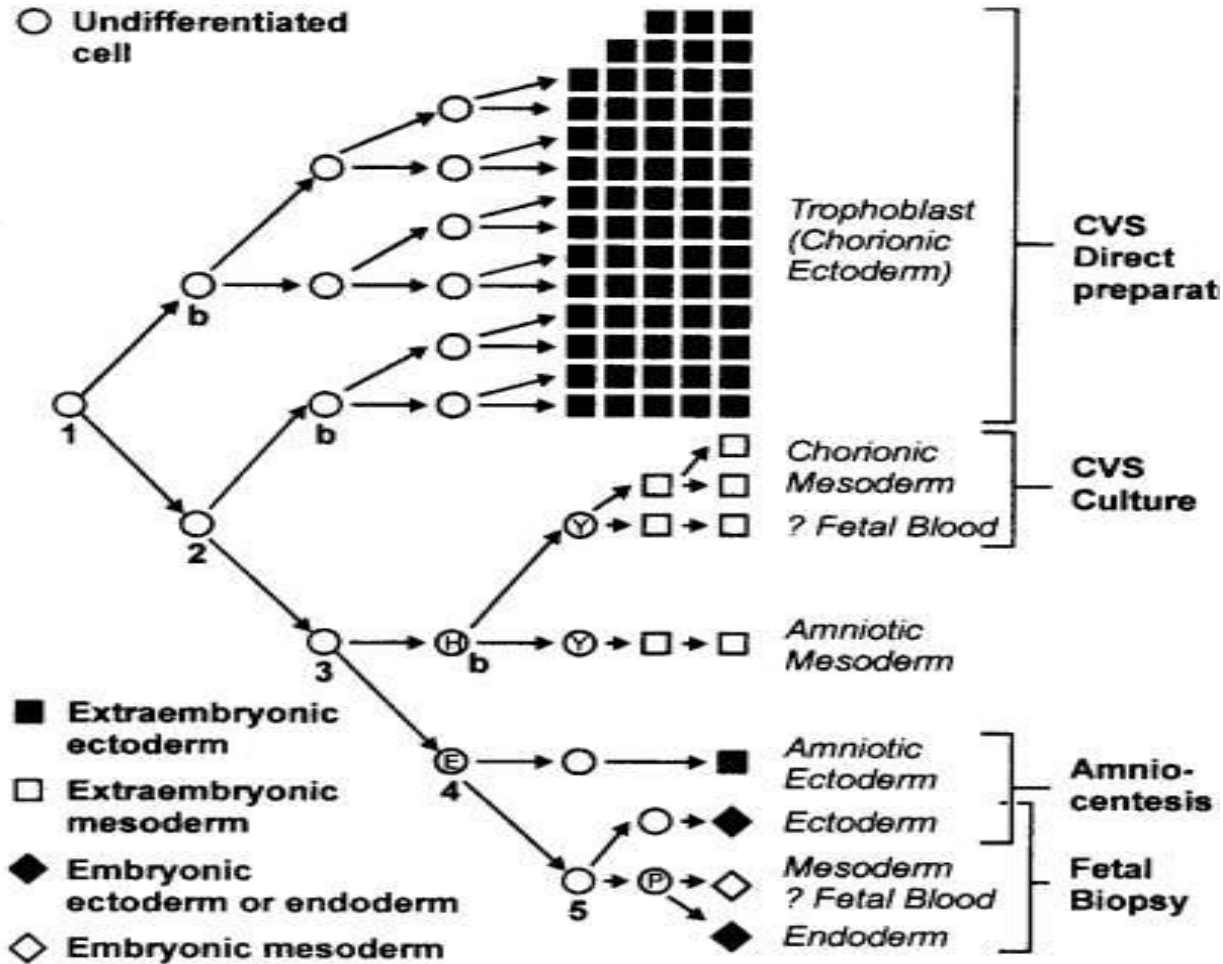
Confined placental Mosaicism

- **Confined placental mosaicism**
 - **An abnormal cell line may only exist in the extra-embryonic tissues of the placenta**
 - **Is encountered at CVS rather than AC**
 - **It is uncommon that mosaicism at CVS reflects a true constitutional mosaicism of the fetus**
 - **More than 50000 procedures (grati et al. 2014)**
 - **In 2,2% of CVS mosaicism was seen -> 0,3% proved to have true fetal mosaicism**

True fetal Mosaicism?

- **Chorion Villi Sampling**
 - **Samples more distantly related from the fetus**
- **Amniocentesis**
 - **Cells closely reflect the true constitution of the fetus**

Embryological Origins



Gardner & Sutherland 3rd Edition 2004